

Ready-to-use BCA Protein Assay Kit

R1491648

Introduction:

The BCA method is one of the most widely used techniques for protein quantification. This kit, based on the bicinchoninic acid (BCA) assay, enables rapid, stable, and sensitive measurement of protein concentrations. Its principle relies on the reduction of Cu^{2+} to Cu^+ by peptide bonds in an alkaline environment, followed by the formation of a purple-colored complex between Cu^+ and BCA. This complex exhibits strong absorbance at 562 nm, proportional to protein concentration. The kit includes a series of ready-to-use protein standard solutions (BSA), eliminating the need for dilution and simplifying the workflow.

Product Component List and Storage Conditions:

| R1491648 | Component | 500T | 500T×5 | Storage |
|-----------|---------------------------------------|--------|----------|---------|
| R1491648A | Reagent A | 100 mL | 100 mL×5 | 2-8°C |
| R1491648B | Reagent B | 3 mL | 3 mL×5 | 2-8°C |
| R1491648C | Ready-to-use BSA standard①(0μg/mL) | 1 mL | 1 mL×5 | -20°C |
| R1491648D | Ready-to-use BSA standard②(125μg/mL) | 1 mL | 1 mL×5 | -20°C |
| R1491648E | Ready-to-use BSA standard③(250μg/mL) | 1 mL | 1 mL×5 | -20°C |
| R1491648F | Ready-to-use BSA standard④(500μg/mL) | 1 mL | 1 mL×5 | -20°C |
| R1491648G | Ready-to-use BSA standard⑤(750μg/mL) | 1 mL | 1 mL×5 | -20°C |
| R1491648H | Ready-to-use BSA standard⑥(1000μg/mL) | 1 mL | 1 mL×5 | -20°C |
| R1491648I | Ready-to-use BSA standard⑦(1500μg/mL) | 1 mL | 1 mL×5 | -20°C |
| R1491648J | Ready-to-use BSA standard⑧(2000μg/mL) | 1 mL | 1 mL×5 | -20°C |

Key Features:

1. Convenient: Ready-to-use standards avoid tedious dilution steps.
2. High Accuracy: Lower coefficient of variation than Coomassie Brilliant Blue assays.
3. Broad Linear Range: Detection range: 20–2000μg/mL.
4. Excellent Compatibility: Tolerates metal ions, reducing agents, chelators, and detergents.

Protocol (Microplate Reader Example):

1. Prepare Working Reagent:
 - 1.1 Calculate the total volume required:
 Working reagent volume = (Number of standard samples + Number of test samples) × Replicates × Volume per well.
 Example: 8 standards + 3 test samples with 3 replicates each at 200μL/well:
 $(8 + 3) \times 3 \times 200\mu\text{L} = 6.6 \text{ mL}$

1.2 Mix Reagent A and Reagent B at a 50:1 ratio to prepare the working reagent. Vortex thoroughly.

Note:

- Prepare extra volume (1–2 wells) to compensate for pipetting errors.
- Freshly prepared working reagent is stable for 24 hours at room temperature (sealed).

2. Quantification Steps:

2.1 Add 20 μ L of each ready-to-use BSA standard (①–⑧) to a 96-well plate (mix thoroughly before use):

| Well | 1 | 2 | 3 | 4 | 5 | 6 | 7 | 8 |
|-------------------|----|-----|-----|-----|-----|------|------|------|
| Standard | ① | ② | ③ | ④ | ⑤ | ⑥ | ⑦ | ⑧ |
| Volume (μ L) | 20 | 20 | 20 | 20 | 20 | 20 | 20 | 20 |
| BSA (μ g/mL) | 0 | 125 | 250 | 500 | 750 | 1000 | 1500 | 2000 |

2.2 Dilute test samples in 1 \times PBS or 0.9% saline (e.g., 2 \times , 4 \times , 8 \times serial dilutions). Add 20 μ L to wells.

2.3 Add 200 μ L working reagent to each well. Mix thoroughly, cover the plate, and incubate at 37°C for 30 min. Cool to room temperature.

Note: Incubation alternatives:

- Room temperature: 2 hours.
- 60°C: 30 minutes.
- Absorbance increases with time/temperature. Extend incubation for low-concentration samples.

2.4 Measure absorbance at 562 nm (or 540–590 nm) using a microplate reader. Subtract the blank control (Standard ① + working reagent) absorbance.

2.5 Plot a standard curve and calculate sample protein concentrations.

Note:

- Exclude obvious outliers.
- Multiply calculated concentrations by dilution factors.
- Use linear regression for computer-based fitting.

Matters needing attention:

1. Compatible with microplate readers (microplate method) or spectrophotometers (cuvette method). For cuvettes, ensure the volume meets the minimum requirement (may reduce total number of assays).
2. If precipitates form during cold/long-term storage, stir or warm to 37°C to dissolve.
3. Always generate a fresh standard curve for each assay to ensure accuracy.
4. For samples with high levels of interfering substances (see Appendix), consider alternative protein quantification methods.
5. Wear a lab coat and disposable gloves during operations.
6. For research use only.